

DNA Sequencing Core Workflow & Customer Interaction

Helaman Escobar





Biostatistics

Cell Imaging/ Fluorescence Microscopy

DNA Sequencing

DNA/Peptide

Electron Microscopy

Flow Cytometry

Genomics

Mass Spectrometry and Proteomics

Microarray

Microarray/Bioinformatics

NMF

Protein Interaction

Small Animal Imaging

Transgenic/Gene-Targeting Mouse Facility

C. elegans 'Worm' Facility

CZAR (zebrafish)

Amaxa nucleofector

nucleofector login request

Core Facility News/Announcements

from Jerry Kaplan, Associate Dean for Research, School of Medicine

- We have recently obtained two new pieces of equipment that may be used by faculty.
- a. The Huntsman General Clinical Research Center (GCRC) in the School of Medicine recently acquired an AUTOPURE automated DNA extraction system. This system is capable of processing up to 96 samples per day and can be programmed to prepare DNA from whole blood, tissue culture cells and buccal swabs.

The GCRC will be accepting applications from the entire University of Utah campus to provide DNA preparation service. Applications in the form of an abbreviated protocol will be available from the GCRC and will be screened for approval by the GCRC Advisory Committee. This service will be free of charge to investigators with approved DNA preparation only protocols. If sample numbers for a single protocol are high, the principal investigator will be asked to cover the cost of reagents.

DNA samples will be verified by gel electrophoresis, A260/280 ratios will be provided for each sample and returned at a standard concentration of 200 ug/ml. Details and protocol forms will be posted soon on the GCRC website http://crc-gw.med.utah.edu/index.cfm.

b. The Health Science Center has purchased four mouse metabolic cages, each one houses one mouse during a typical experiment. These cages allow one to measure oxygen consumption, CO2 evolution, heat generation, locomoter activity, food intake, water intake and related parameters on normally behaving mice in real time. This equipment will be housed in the Department of Biochemistry. For additional information please contact Jared Rutter, Ph.D., Office: 581-3340, Lab: 581-8718 Rutter@biochem.utah.edu





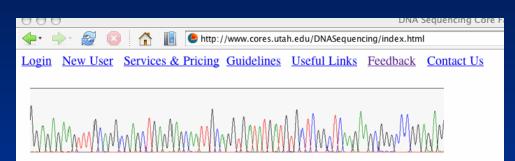
Research Resources

Grant information, training grants that exist on campus, location of supply centers/freezer programs, etc.

- Biostatistics
- Cell Imaging
- DNA Sequencing
- & Genomics
- Electron Microscopy
- Flow Cytometry
- Proteomics
- Microarray
- Bioinformatics
- •NMR
- Protein Interaction
- •Small Animal Imaging
- •Transgenic Mouse
- •C. elegans
- Zebrafish
- Metabolomics

www.cores.utah.edu





DNA Sequencing Core Facility

The DNA Sequencing Core Facility at the University of Utah Health Science Center uses state of the art fluorescent DNA sequencing methods to produce high quality DNA sequencing at competitive price to the University community and off-campus users. The facility is equipped with one ABI3730 96-capillary sequencer and one ABI3100 16-capillary sequencer. The average turnaround time is two working days.

Lastest News

01/12/06	The Core is now hosting a Freezer Program for Applied Biosystems Lab Consumables. Get it cheaper, get it faster and pay no shipping (if we don't have it in stock). Download the order form and item list here (pdf file) or call us for more info.
04/22/05	Sequencher TM 4.5 for Mac and PC is now available.
01/13/05	You can now send us feedback on any issue concerning our services through this web site. Click on the <u>feedback</u> link above.

How can I open and edit my sequence files? Click on the link below: Sequencher TM 4.5 for MacOS 9.2 or 10.2 and higher; includes OS X users









Chris Walls

Mike Klein



Equipment



ABI 7900 HT



Robotics



ABI 3130xI



Biomek FX



ABI 3730xl



Other



Lab Profile – DNA Sequencing

- Not subsidized
- 210 Pls and 550 users (techs + students)
- 5 pick-up locations on campus
- 100K sequencing samples (cycle seq)
- 30K samples for re-sequencing projects
- 40% of work requests 4 samples or less



Basic Workflow

- 1. Re-array all samples into 96-well plates
- 2. Transfer to 384-wells before cycling
- 3. EtOH Precipitation
- 4. Electrophoresis in formamide
- Review data using LIMS + SeqA or SeqScape



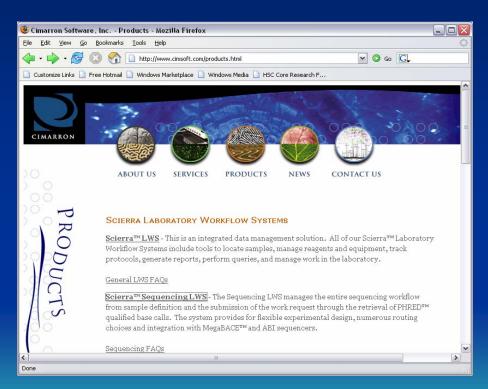
Workflow Highlights

- LIMS
- Use robotics Velocity11 Vprep Station
- We pre-mix and aliquot BD 3.1
 - 1/32 BD dilution
- Same plate from reaction to instrument
- 5 ul rxns:
 - 0.5 pmoles of primer
 - 5 -10 ug PCR or 100-150 ug of plasmic
 - Very long array life 700 to 1100 runs
- 2% dropout rate





Scierra from Cimarron



- Flexibility
- Oracle Database
- Other Core modules
- Tracking every process step
- Barcode enabled
- Flag problems
- Reassign samples
- Built-in inventory
- Records protocols and processes

www.cimsoft.com



Do I need a LIMS?

- Increase your productivity, minimized human error, free time and please your customers
- More than 15 20K samples
- Out-of-the-box decide what features are must
- If building your own:
 - Make sure you have a Requirements Specification
 Document
 - Hire professionals



The Core Lab Business Predicament

- Quality
 - Seq Data quality
 - Customer service/support
- Turn over time
- Price

What should be my focus?



Business Approach

- Assembly line
- Adhere to a Schedule
 - Clear time deadlines
- Consistent Processing
 - Detailed protocols
 - Streamline your format
 - Delegate specific tasks to technicians

Pros:

Predictability
Minimize human error
Less stress
Consistent results
Time saving

Cons:

Takes a certain type of staff Goes against academic culture Extra consumables cost



Turnover and Quality

- We check most sequences individually
- Redo dropouts automatically
- Contact users having systematic problems
- 25 hrs turnover time
- Perform surveys
- Hold Lab Open houses and sponsor seminars
- Common lab area

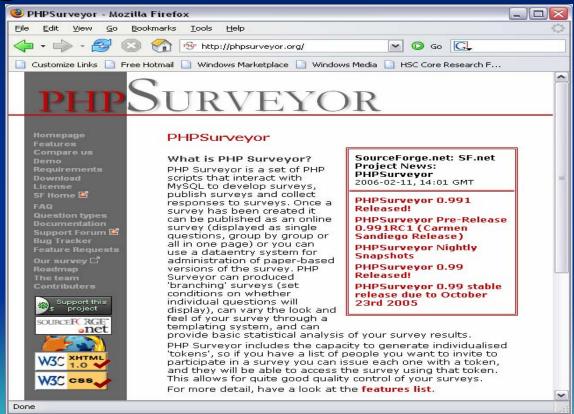


How to get to your Customers

- Redundancy
- Redundancy
- Redundancy



Surveys



http://phpsurveyor.org



Survey Caveats

- Make them short!!
- Ask meaningful questions
- Do not do them too often!
- Act on them quickly!!
- Share the data
- Personally contact those who raised issues and gave feedback



Case Study: Post-sequencing Analyzes

- 26 questions most of them multiple choice
- 4 min to complete
- Opened for 2 weeks
- Major questions:
 - Sequence software
 - Computer platform
 - Expectations from the Core



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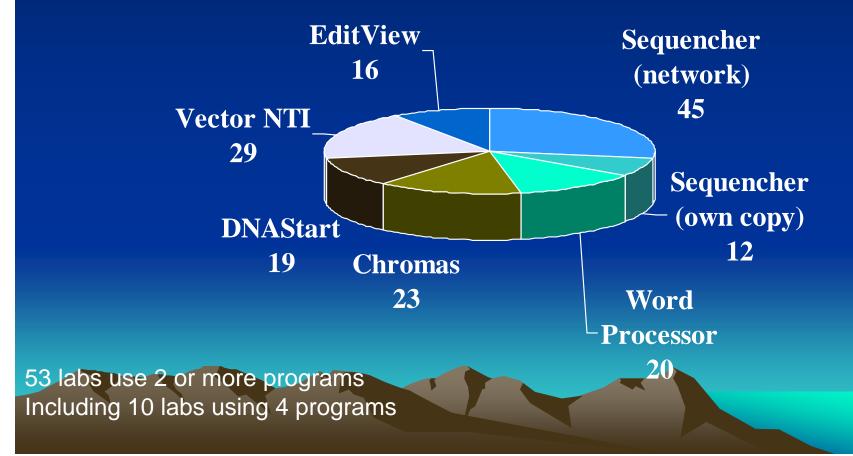
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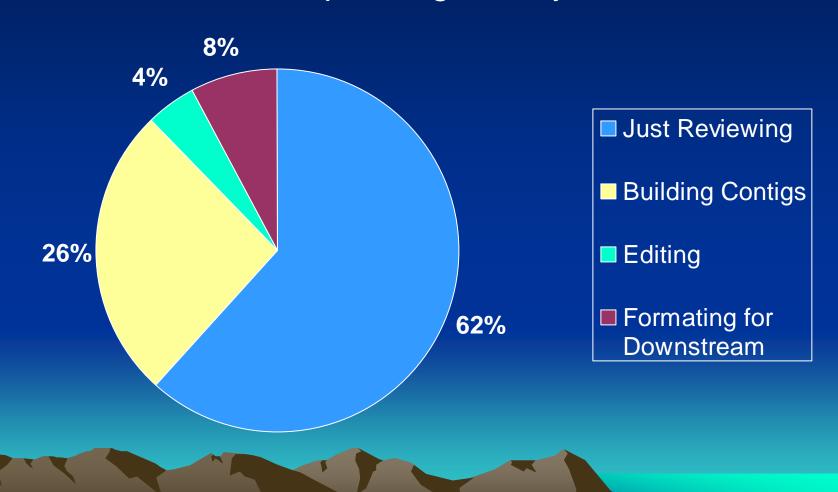
Most Comon Sequence Editors

N = 124





Post Sequencing Activity





Feedback

- Software training session for Sequencher
- A Sequencing Troubleshooting Guide
- Forum of a Blog were users can exchange information
- Training on other Sequence Editors
- Phred files posted.



List of Common Editors

- Sequencher
- Chromas
- MacVector
- DNA Stars
- DNAman
- DNA Strider
- Vector NTI

- BioEdit
- 4Peaks
- Ape
- Multalin (web)
- ClustalW
- GCG
- Staden Pkg
- NCBI



Survey Conclusions

- I don't need more licenses for Sequencher
- I don't need to upgrade Sequencher
- I need to provide training for downstream software
- Software they use might be influencing their Core perceived quality



Review

- Run your Core like a small business
- Decide the focus for your business
- Be consistent with your workflow and sample processing
- Develop good communication with your customers
- Enjoy It